

Reducing human exposure to aflatoxin through the use of clay: A review

T. D. Phillips , E. Afriyie-Gyawu , J. Williams , H. Huebner , N.-A. Ankrah , D. Ofori-Adjei , P. Jolly , N. Johnson , J. Taylor , A. Marroquin-Cardona , L. Xu , L. Tang & J.-S. Wang

To cite this article: T. D. Phillips , E. Afriyie-Gyawu , J. Williams , H. Huebner , N.-A. Ankrah , D. Ofori-Adjei , P. Jolly , N. Johnson , J. Taylor , A. Marroquin-Cardona , L. Xu , L. Tang & J.-S. Wang (2008) Reducing human exposure to aflatoxin through the use of clay: A review, Food Additives and Contaminants, 25:2, 134-145, DOI: [10.1080/02652030701567467](https://doi.org/10.1080/02652030701567467)

To link to this article: <https://doi.org/10.1080/02652030701567467>



Published online: 20 Feb 2008.



Submit your article to this journal [↗](#)



Article views: 2261



View related articles [↗](#)



Citing articles: 79 View citing articles [↗](#)

Reducing human exposure to aflatoxin through the use of clay: A review

T. D. PHILLIPS¹, E. AFRIYIE-GYAWU¹, J. WILLIAMS², H. HUEBNER¹,
N.-A. ANKRAH³, D. OFORI-ADJEI³, P. JOLLY⁴, N. JOHNSON¹, J. TAYLOR¹,
A. MARROQUIN-CARDONA¹, L. XU⁵, L. TANG⁵, & J.-S. WANG⁵

¹Department of VIBS-MS 4458, Texas A&M University, Veterinary Integrative Biosciences, College of Veterinary Medicine, College Station, TX 77843, USA, ²Peanut Collaborative Research Support Program, University of Georgia, Griffin, GA, USA, ³School of Public Health, Noguchi Memorial Institute for Medical Research, University of Ghana, Legon, Accra, Ghana, ⁴Epidemiology, University of Alabama at Birmingham, Birmingham, AL, USA, and ⁵Institute of Environmental/Human Health, Texas Tech University, Box 41163, TTU/TIEHH, Lubbock, TX 79409-1163, USA

(Received 18 May 2007; accepted 10 July 2007)

Abstract

Innovative sorption strategies for the detoxification of aflatoxins have been developed. NovaSil clay (NS) has been shown to prevent aflatoxicosis in a variety of animals when included in their diet. Results have shown that NS clay binds aflatoxins with high affinity and high capacity in the gastrointestinal tract, resulting in a notable reduction in the bioavailability of these toxins without interfering with the utilization of vitamins and other micronutrients. This strategy is being evaluated as a potential remedy for acute aflatoxicosis, and as a sustainable human intervention for aflatoxins via the diet. Phase I and II clinical trials confirmed the apparent safety of NS for further study in humans. A recent study in Ghanaians at high risk for aflatoxicosis has indicated that NS (at a dose level of 0.25%) is effective in decreasing biomarkers of aflatoxin exposure and does not interfere with the levels of serum vitamins A and E, and iron and zinc. In summary, enterosorption strategies/therapies based on NS clay are promising for the management of aflatoxins and as a sustainable public health intervention. The NS clay remedy is novel, inexpensive and easily disseminated. Based on the present research, aflatoxin sequestering clays should be rigorously evaluated *in vitro* and *in vivo*, and should meet the following criteria: (1) favourable thermodynamic characteristics of mycotoxin sorption, (2) tolerable levels of priority metals, dioxins/furans and other hazardous contaminants, (3) safety and efficacy in multiple animal species, (4) safety and efficacy in long-term studies, and (5) negligible interactions with vitamins, iron and zinc and other micronutrients.

Keywords: *NovaSil clay, mycotoxins, aflatoxins, aflatoxin-binding agent, aflatoxin sorbent, aflatoxin-sequestering agent, clinical trial, Ghana*

Introduction

Historical perspective on moulds

Moulds (and their metabolic by-products) can be beneficial, as well as harmful, to humans and animals. Moulds have been used since ancient times in the production of various foods including cheese and salami and in the fermentation of beer and wine (Peraica et al. 1999). The secondary metabolites from these same types of moulds have been used as very effective antibiotics for the treatment of disease and as drugs for other important

medicinal purposes. For example, the Chinese used moulds for obstetrical purposes nearly 5000 years ago (Hesseltine 1979). Unlike bacterial toxins, the mycotoxins are not protein in nature, but consist of highly diverse organic structures characterized by a variety of heteroatom-containing functional groups. Many of these potent organic chemicals, though invisible to the naked eye, can be found in mould-contaminated food sources and may be harmful if ingested in high enough quantities or over a long enough period of time. Fortunately, the toxicity of these compounds is dose related, and their levels in

foods and feeds are typically low to non-detectable. However, during extended periods of drought the production of certain hazardous mycotoxins can be unavoidable and may result in contaminated food and feed products that present significant health risks to man and animals (Phillips et al. 2002, 2006; Huebner et al. 2004; Williams et al. 2004).

Mouldy food poisoning and human disease

Although the term ‘mycotoxin’ was not commonly used until the mid-20th century, earlier records suggest that mycotoxin contamination of food and major outbreaks of disease associated with the consumption of mouldy food have occurred frequently throughout history. A variety of toxic effects of moulds can be traced to very early civilizations, including the Chinese almost 5000 years ago (Ramsbottom 1953; Van Rensburg and Altenkirk 1974). Of the more than 300 mycotoxins that have been identified and chemically characterized, many have been found as contaminants of food and have been linked to the aetiology of disease in humans (and animals). Of these, the aflatoxins have been extensively studied due to their frequent occurrence in foods (especially in developing countries) and their mutagenicity and carcinogenicity (Wogan 1992; Wild and Hall 2000; Wild and Turner 2002).

Aflatoxins

Discovery

There was a lack of understanding of the consequences of aflatoxin exposure on human and animal health, until the early 1960s, when mouldy feed was associated with the loss of thousands of young turkeys in the UK. In this incident, the affected animals showed signs of severe liver necrosis as well as fatty degeneration, fibrosis and extensive bile-duct hyperplasia (Siller and Ostler 1961). Upon investigation it was discovered that the Turkeys had been fed Brazilian peanut meal containing the mould *Aspergillus flavus* along with four metabolic by-products, namely aflatoxins B₁ (AfB₁) (Figure 1), B₂ (AfB₂), G₁ (AfG₁) and G₂ (AfG₂) (Asao et al. 1963). For confirmation the same symptoms were produced in a variety of other species, including ducklings (Sargeant et al. 1961) and rats (Lancaster et al. 1961) following ingestion of the contaminated peanut meal. The dramatic effects of the aflatoxins resulted in significant scientific interest in delineating the chemical structures and toxicological properties of aflatoxins (and other mycotoxins).

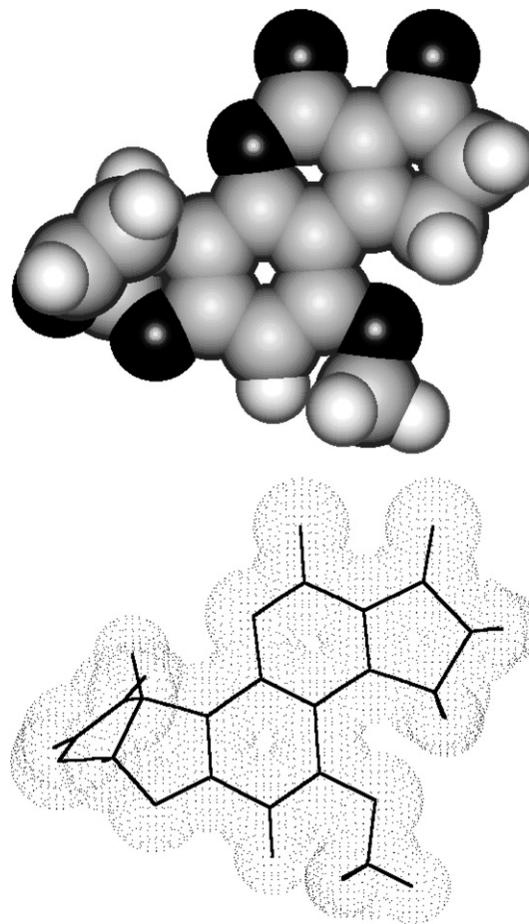


Figure 1. Molecular model of aflatoxin B₁ showing the spatial arrangement of atoms. The molecule is planar, except for the terminal furan which is kinked in the *cis* configuration (coming out of the page). White, hydrogen; dark grey, oxygen; grey, carbon.

Toxic effects

The first observations regarding the toxicity of aflatoxins were recorded during the Turkey X incident in the UK in 1960. In these birds, acute necrosis and bile duct proliferation of the liver was observed (Lancaster et al. 1961). Soon after the same types of symptoms were reported in ducklings and pheasants (Sargeant et al. 1961). A number of acute dosing studies have been performed with AfB₁ to determine LD₅₀ values for a wide range of animals (Council for Agricultural Science and Technology (CAST) 1989). While AfB₁ was found to be toxic for many species, some animals were identified as highly sensitive, including duckling, rabbit, and rainbow trout (Muller et al. 1970). Other common symptoms of acute poisoning by AfB₁ include depression and anorexia, as seen in the recent contamination of pet food in South Texas (Garland and Reagor 2001) and South Carolina (Lang 2006). Chronic toxicity studies have also been conducted with lower levels

of aflatoxin exposure. One of the major effects is a general reduction in weight gain for a variety of production animals, including pigs, cattle and poultry. Also, milk production in dairy animals can be decreased in the presence of aflatoxin-contaminated food (CAST 1989) and the milk carries a metabolite of known as AfM₁. This chemical is highly regulated because infants and young children may consume large quantities of milk, and the young of all species are more susceptible (than adults) to the effects of aflatoxins (Leeson et al. 1995).

The main effect of chronic exposure to aflatoxin in humans is hepatocellular carcinoma (HCC). While the incidence of this disease is low in the USA, in parts of Africa and Asia these numbers can be very high (Groopman et al. 1988). Epidemiological studies have shown a positive correlation between the intake of aflatoxin and liver cancer among African and Asian populations. For example, a study by Bulatao-Jayme et al. (1982) showed that the relative risk for liver cancer, when consuming a large level of aflatoxin with a minor amount of alcohol, was 17.5 as compared with a relative risk of 3.9 when alcohol consumption was heavy and aflatoxin intake was light. A diet dependent on foods such as cassava, peanuts, sweet potato and corn increases the likelihood of consuming mould-infested food. A serious confounder in these studies was the high rate of hepatitis B virus (HBV) in these populations. The potential interaction of HBV with aflatoxin is not fully understood (Groopman et al. 1988). The carcinogenic potency of AFB₁ in individuals positive for hepatitis B virus (HBV) surface antigen (HBsAg) has been reported to be considerably higher compared with individuals who are negative for HBsAg. Most of the epidemiological data are from geographical areas where both the prevalence of HBsAg + individuals and aflatoxins are high; the relationship between these risk factors in areas of low aflatoxin contamination and low HBV prevalence is unclear (World Health Organization (WHO) 1998).

Biochemical mode of action

AfB₁ is a direct-acting mutagen and identification of a DNA adduct was made by Essigmann et al. (1977). It was shown that the 8,9 vinyl ether group is transformed to an epoxide through cytochrome P450-mediated oxidation; carbon 8 on aflatoxin reacts with guanine at the N⁷ position to form an 8,9-dihydro-8-(N⁷-guanyl)-9-hydroxy-AfB₁ adduct (Figure 2). Studies have utilized nuclear magnetic resonance (NMR) to characterize the intercalation and adduct formation of AfB₁ with two oligodeoxynucleotide sequences

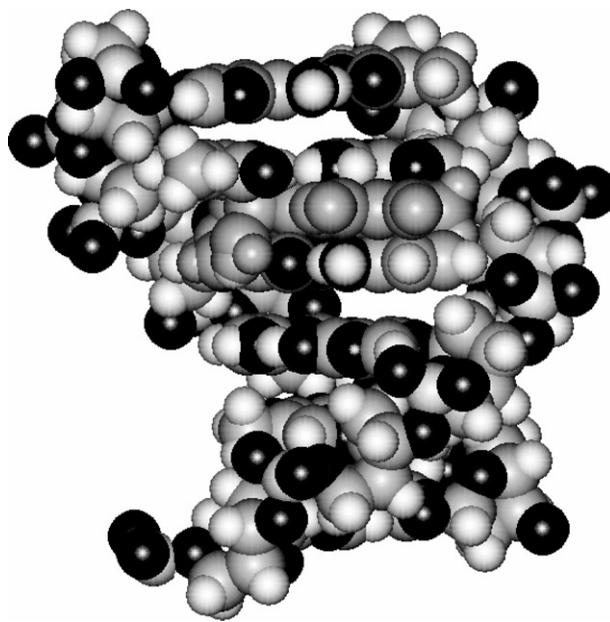


Figure 2. Molecular model showing AfB₁ (in medium grey) intercalated between the strands of DNA.

[d(ATC^{AFB}GAT).d(ATCGAT) and d(AT^{AFB}G-CAT)₂] (Gopalakrishnan et al. 1990). Importantly, AfB₁ has been shown to disrupt several genes involved in the growth of cancer. When administered to rats, it was demonstrated that the resulting liver tumours contained an activated form of a proto-oncogene of the c-Ki-ras family. In addition, the hepatocellular tumours of AfB₁-treated rats showed high expression of the proto-oncogenes c-H-ras and c-myc (McMahon et al. 1986, 1987). Proto-oncogenes stimulate growth in the normal cell; when mutated, loss of function leads to rapid growth characteristic of cancer. Tumour suppressor genes provide a complement to this system, keeping growth in check in the normal cell. AfB₁ has been shown to produce a G → T transversion in the p53 tumour suppressor gene in human hepatocytes (Aguilar et al. 1993). In addition to DNA and RNA damage, AfB₁ has also been shown to interact with RNA and intercellular proteins. Interactions with protein (Sabbioni et al. 1987; Guengerich et al. 2002a, 2002b) may explain some of the non-carcinogenic effects following exposure to aflatoxin (Eaton et al. 1994).

Consequences of exposure

AfB₁ has been described as a human carcinogen (Group 1A) and implicated in HCC (International Agency for Research on Cancer (IARC) 1976, 1987, 1993). Also, studies suggest that AFs impair the cellular and humoral immune system in animals, and that low-level exposure to these toxins can cause immunosuppression and increased susceptibility to

disease (Rodricks and Stoloff 1977; Miller et al. 1978; Richard et al. 1978; Peska and Bondy 1994; Hinton et al. 2003). A study by Turner et al. (2003) in Gambian children showed evidence that secretory IgA in saliva may be reduced from dietary aflatoxin exposure. This was the first report of immunosuppression in humans associated with AF biomarker measures. Jiang et al. (2005) recently confirmed this finding in adult humans in Ghana and showed a significant correlation between aflatoxin exposure and suppression of the immune system. Other consequences of dietary exposure to aflatoxins include adverse effects on growth and antinutritional effects in animals. For example, AFB₁ has been shown to reduce hepatic vitamin A significantly in a variety of animals, including chickens (CAST 1989; Pimpukdee et al. 2004; Williams et al. 2004). Importantly, Gong et al. (2002, 2004) and Turner et al. (2007) reported an association between biomarkers of aflatoxin exposure and growth impairment in children in West Africa.

Although many countries have regulatory limits for aflatoxins in foods/feeds, outbreaks of poisoning frequently occur. A recent outbreak of aflatoxin poisoning in Kenya resulted in a 39% case fatality rate and was linked to consumption of foods containing toxin levels as high as 8000 ng g⁻¹ (Centers for Disease Control and Prevention (CDC) 2004). Drought stress exacerbates fungal infection, thus enhancing production of the aflatoxins. This is especially true between a latitude of 40°N and 40°S of the equator, a hot zone which encompasses many developing countries where aflatoxins in the diet of humans and animals are largely uncontrolled (Williams et al. 2004). The poorest people who are most likely to consume foods contaminated with aflatoxins suffer the most severe effects, including disease and even death following acute exposure (Lewis et al. 2005). Additionally, it is estimated that 80% of all HCC cases occur in developing countries (Wild and Hall 2000). Thus, feasible interventions and therapies to diminish human and animal exposure to aflatoxins are imperative; dietary calcium montmorillonite clay, used as an aflatoxin enterosorbent, may provide a practical, cost-effective, and sustainable solution to the problem.

Intervention approaches

Chemopreventive agents

Because avoiding consumption of aflatoxin-contaminated foods for many is simply not feasible, effective means for reducing dietary exposure to aflatoxins are highly desirable (Phillips et al. 2006). Chemoprevention is one strategy used to solve the

problem in high-risk populations. This involves the use of natural or synthetic agents to block, retard, reverse or modulate the carcinogenic process (Gupta and DuBois 2001; Sporn and Suh 2002). A variety of chemopreventive agents exist as natural constituents in the human diet; many of these include phytochemicals derived from various sources. Although efficacious against a wide range of carcinogens, most of these compounds occur at very low levels in a nutritionally balanced diet and are poorly absorbed in the gastrointestinal tract (Hayatsu et al. 1988; Dragsted et al. 1993). The chemopreventive agent oltipraz, an antischistosomal drug, has been evaluated for use in humans exposed to dietary aflatoxins in China (Kensler et al. 1999; Wang et al. 1999). In clinical trials oltipraz, when administered to individuals exposed to dietary aflatoxins, increased the level of glutathione *S*-transferase-mediated conjugation of aflatoxin 8,9-epoxide, but also inhibited cytochrome P450 1A2 activity, a key enzyme that activates aflatoxin to the reactive epoxide (Wang et al. 1996, 1999; Kensler et al. 1999). Oltipraz may also inhibit hepatitis B virus (HBV) transcription through elevation of p53 providing an additional contribution to HCC chemoprevention (Chi et al. 1998). Chlorophyllins are natural occurring constituents of the human diet that have been shown to be effective anticarcinogens in several animal models (Dashwood et al. 1998). They are hypothesized to act as interceptor molecules by binding with carcinogens, such as AfB₁, thereby diminishing bioavailability by impeding their absorption. (Breinholt et al. 1995). In a 4-month clinical trial in China, consumption of 100 mg of chlorophyllin at each meal led to an overall 55% reduction in median urinary levels of aflatoxin-N⁷-guanine adducts versus the placebo (Egner et al. 2001). Application of these compounds in humans would require careful evaluation including long-term effects of enzyme modulation and potential interferences with the uptake of essential nutrients from the diet. Green tea-derived polyphenols, which are highly effective agents against cancer in various animal models, are also being considered as possible interventions for populations at high risk for HCC. Research has indicated that green tea inhibits the initiation of AfB₁-induced hepatocarcinogenesis in the rat by modulating metabolism of AfB₁ (Qin et al. 1997). Administration of green tea (3% in water) prevented hepatic focal lesion growth induced by dieldrin in B6C3F1 mice (Klaunig and Kamendulis 1999). In humans, inverse associations between the level of green-tea consumption and the risk of development and/or time of cancer onset have also been observed (Nakachi et al. 2000; Fujiki et al. 2002).

Interventions that reduce the dose of aflatoxins from contaminated foods

Food surveillance. Surveillance and subsequent regulation of susceptible commodities, such as groundnuts and maize for aflatoxins and other mycotoxins, are routinely used as a primary intervention to safeguard the health of consumers as well as the economic interests of producers and traders in various countries. These surveillance data are frequently used to establish regulatory guidelines that define the limits of aflatoxins and other mycotoxins in foods. However, in many developing countries, these guidelines are not adequately enforced and result in populations at high risk for aflatoxicosis, i.e. recent outbreak of acute aflatoxin poisoning in Kenya (CDC 2004; Lewis et al. 2005).

Community education. One of the most practical and fundamental interventions at the subsistence-farm level in developing countries, is the use of low-technology approaches, such as community education on food handling and storage, as described by Turner et al. (2005). These primary approaches have been shown to reduce significantly the level of aflatoxin contamination in post-harvest foods and associated exposure in human populations at high risk for aflatoxicosis.

Aflatoxin enterosorption (NovaSil clay). Another strategy for reducing food-borne exposure to mycotoxins is the inclusion of various binding agents or sorbents in the diet. Many of these binding agents are purported to prevent the deleterious effects of diverse mycotoxins in a variety of animals (primarily poultry and swine). As early as 1979, adsorbent clay minerals were reported to bind AfB₁ in liquids (Masimango et al. 1979). Additionally, bleaching clays used to process canola oil were found to lessen the effects of T-2 toxin (Carson and Smith 1983; Smith 1984). The dietary consumption of earth (i.e. geophagy) has been observed for centuries and across all continents in both humans and animals (Carretero 2002). Clay eating has been recorded from traditional human societies and is considered 'culturally acceptable' in many African countries and China (Johns and Duquette 1991; Diamond 1999). A practical approach of current interest for the prevention of aflatoxicoses is the incorporation of non-nutritive clay minerals in contaminated food/feed to sorb aflatoxins in the stomach and intestinal tract, thus reducing toxin bioavailability and distribution to the blood, liver and other target organs (Phillips et al. 1995, 2002, 2006; Phillips 1999). Using multiple animal models, the present authors' laboratory has shown that NovaSilTM (NS) clay,

a calcium montmorillonite, can prevent the adverse effects of exposure to dietary aflatoxins.

Initially, NS (which was referred to as HSCAS in the early literature), was sold as an anticaking additive for animal feeds. It was reported to sorb aflatoxin B₁ with high affinity and high capacity in aqueous solutions and was shown to rescue broiler and Leghorn chicks from the toxic effects of 7500 ppb aflatoxin in the diet (Phillips et al. 1987, 1988, 2006). In subsequent studies, NS and other similar montmorillonite and smectite clays have been reported to protect against aflatoxin toxicity in a variety of young animals including rodents, chicks, turkey poults, ducklings, lambs, pigs, mink and trout (Phillips 1999; Phillips et al. 1990, 1991, 1994, 1995; Colvin et al. 1989; Bonna et al. 1991; Harvey et al. 1991a, 1991b; 1993, 1994; Voss et al. 1993; Kubena et al. 1990a, 1990b, 1991, 1993; Ledoux et al. 1999; Smith et al. 1994; Marquez and Hernandez 1995; Cerdchai et al. 1990; Lindemann et al. 1993; Abdel-Wahhab et al. 1998; Nahm 1995; Jayaprakash et al. 1992; Ellis et al. 2000). In studies using radiolabelled aflatoxins, NS clay has also been shown to decrease the bioavailability of aflatoxins and reduce aflatoxin residues in poultry (Davidson et al. 1987; Jayaprakash et al. 1992), rats (Sarr et al. 1995; Mayura et al. 1998) and pigs (Beaver et al. 1990). Aflatoxin M₁ levels in milk from lactating dairy cattle and goats were also decreased when NS was included in the diet (Ellis et al. 1990; Harvey et al. 1991b; Smith et al. 1994).

Mechanisms of aflatoxin sorption to NS. The suggested mechanism of AfB₁ sorption by NS is an electron donor acceptor (EDA) mechanism. The platelets of NS clay are negatively charged due to isomorphic substitution, and thus they attract positively charged ions to balance this charge. Compounds with areas of electron deficiencies (partial positive areas) can also be attracted to the platelets (Haderlein et al. 1996). The carbons comprising the dicarbonyl system in aflatoxins are partially positive and have been shown to be essential to the adsorption process. AfB₁ is planar with the exception of the terminal furan (Figure 1). The importance of the spatial orientation of AfB₁ was demonstrated when stereochemical differences of some aflatoxin analogues resulted in significant effects on the tightness of binding. These results also suggested that the sorption of aflatoxin onto NS may favour an orientation where the furan is aligned away from the surface. Adsorption isotherms on heat-collapsed NS have demonstrated that the interlamellar region of NS is the primary site of binding with external surfaces accounting for only minor sorptions of aflatoxins. Based on the

thermodynamics, AFB₁ binds strongly to NS, exhibited by an estimated heat of sorption (enthalpy) of -50 KJ mol^{-1} . Interference from compounds with stereochemical restrictive groups could also play an important role in the adsorption process. For the analogues that contain functional groups that make them larger than AFB₁, their insertion, docking and adsorption at surfaces in the interlamellar channel might be restricted. Our results also indicate a good correlation between the magnitude of partial positive charges on carbons C₁₁ and C₁ of the β -dicarbonyl system and the strength of adsorption of planar ligands, suggesting an EDA mechanism with the surface of the clay. Other mechanisms of AFB₁ sorption to NS surfaces involve the potential chelation of interlayer cations (especially Ca²⁺) and various edge-site metals (Grant 1998; Phillips 1999; Phillips et al. 2002, 2006).

Selectivity of NS clay. Research has demonstrated that NS clay has a notable preference (and capacity) for aflatoxins. NS at a level of 0.5% w/w in the diet of poultry did not impair phytate or inorganic phosphorous utilization (Chung and Baker 1990). In other studies in poultry, the addition of NS at concentrations of 0.5% (which is recommended for anticaking in feeds), did not impair the utilization of riboflavin, vitamin A, manganese, or zinc (Chung et al. 1990). NS (0.5% w/w) was also shown to protect young chickens from aflatoxin levels as high as 7500 ppb; these levels are not likely to be found in human food; although, the recent exposure of humans in Kenya was linked to toxin levels as high as 8000 ppb. While clay-based interventions are clearly effective for aflatoxins, the same effectiveness has not been demonstrated for other mycotoxins. Importantly, unmodified NS clays have not been shown to strongly bind other structurally diverse mycotoxins, e.g. zearalenone, deoxynivalenol, T-2 toxin, ochratoxin A, cyclopiazonic acid, ergotamine, and fumonisins, nor do they significantly prevent the adverse effects of these mycotoxins when included in the diet of animals. For example, in enterosorbent studies in poultry with mycotoxins other than aflatoxin, the inclusion of NS clay in the diet did not prevent the adverse effects of cyclopiazonic acid (Dwyer et al. 1997), T-2 toxin (Kubena et al. 1990a), diacetoxyscirpenol (Kubena et al. 1993), ochratoxin A (Huff et al. 1992), and fumonisins (Lemke 2000). The inclusion of clay in the zearalenone-contaminated diets of mink alleviated some fetotoxicity, but did not reduce the hyper-estrogenic effects (Bursian et al. 1992). Also the average daily weight gain was unchanged in pigs exposed to deoxynivalenol when clay was added to the diet at 0.5 and 1.0% w/w. The only effective

method for decreasing deoxynivalenol toxicity was dilution of the contaminated maize (Patterson and Young 1993). Although *in vitro* tests showed potential for protection of ergotamine toxicity with NS (Chestnut et al. 1992; Huebner et al. 1999), NS at levels of 2.0% w/w did not protect rats or sheep from fescue toxicosis. NS's selectivity was further demonstrated in our laboratory in studies involving nanostructured thin films of NS on quartz that were used as affinity probes for aflatoxins in contaminated media. Our findings show that this composite exhibited comparable selectivity to the Aflatest affinity column from Vicam (Huebner and Phillips 2003; Huebner et al. 2004).

Long-term safety study in rats. In earlier studies in animals, no observable adverse effects from NS were reported following ingestion of doses up to 2.0% w/w in the diet. For example, Sprague-Dawley rats that ingested NS clay at dietary concentrations as high as 2% throughout pregnancy, did not show significant trace metal bioavailability in a variety of tissues and showed neither maternal nor foetal toxicity (Wiles et al. 2004). Since most of our preliminary work was based on short-term exposures not greater than 6 weeks in duration, a long-term exposure was warranted to establish the safety of NS further. Before an adverse events/dosimetry trial in humans, a rodent model was used to evaluate the relative safety of chronic exposure to NS clay in the diet. Male and female Sprague-Dawley rats were fed rations containing 0, 0.25, 0.5, 1.0, and 2.0% levels of NS clay *ad libitum* over a period of 6.5 months. No morbidity or mortality was observed in the animals throughout the study duration. The results of this study indicated that rats treated with 0.25–2% NS clay in the diet did not exhibit dose-dependent or NS-related adverse effects on body weight gains, feed conversion ratios, relative organ weights, gross anatomy and histological appearance of major organs, haematology, and serum biochemistry parameters. Additionally, levels of selected nutrients including vitamins A and E, Fe, and Zn were unaffected (Afriyie-Gyawu et al. 2005). Given the safety and efficacy of NS, as demonstrated in a variety of animal models, it was hypothesized that NS-based interventions might be beneficial for the treatment of humans who are at high risk for aflatoxicosis (Phillips et al. 2006).

Adverse events/dosimetry trial with NS in humans. As a precursor to a phase IIa clinical intervention trial with NS in Ghana, a short-term (2-week) safety evaluation of NS was carried out at Texas Tech University in 50 healthy adults (Wang et al. 2005). The overall design followed the guidelines

for a randomized and double-blind phase I clinical trial. This study was conducted: (1) to evaluate the short-term safety and tolerance of NS capsules in normal human subjects; and (2) to establish optimal protocols for human intervention studies. NS capsules were produced under sterile conditions using US Good Manufacturing Practices. Also, the NS was sterilized at 121°C before encapsulation. Preceding the chronic animal and short-term human studies, NS was analysed for concentrations of various environmental contaminants, including priority toxic metals and dioxins/furans to ensure compliance with federal and international standards. A total of 50 adults, ages 20–45, who met the recruiting criteria were randomly divided into two study groups. The high-dose group (HD) took three capsules of NS three times a day (a total of 3.0 g), and the low dose group (LD) took three capsules of NS three times a day (a total of 1.5 g). All capsules were of the same colour and size. The two dose levels were extrapolated from previously published dosimetry data from animal studies (Phillips 1999; Phillips et al. 2002, 2004; Afriyie-Gyawu et al. 2005). After 14 days of capsule ingestion, NS (up to 3.0 g day⁻¹) was considered safe for further human studies based on physical examination, biochemistry and haematology results. Concentrations of the standard parameters analysed after the trial were statistically similar to those levels determined before trial. Also, no significant difference was observed for any reported adverse symptoms between LD and HD groups. This study confirms the selectivity of NS clay for aflatoxins, in that no statistical differences were observed in the levels of serum vitamins A and E, and iron and zinc in the participants after 2 weeks of NS ingestion. This evidence further confirms that NS demonstrates binding specificity for aflatoxins and lack of interaction with vitamins A and E. The adverse events trial provided the basis for the phase IIa human intervention study at the Ejura-Sekyedumase district (ESD) of the Ashanti region of Ghana, West Africa.

Screening of clays in Ghana for aflatoxin binders. Before the initiation of a 3-month clinical intervention trial in Ghana, 73 ‘edible’ clays from the marketplace at ESD and 11 clays used in the ceramic industry from other locations in Ghana were tested for aflatoxin sorption using isothermal analyses in our laboratory. Our rationale for screening clays near the study site in Ghana was to identify those that were similar to NS clay for subsequent human studies. It was hypothesized that study participants could be categorized as geophagic or non-geophagic in the context of aflatoxin exposure. However, upon analysis of 84 samples from different geographic

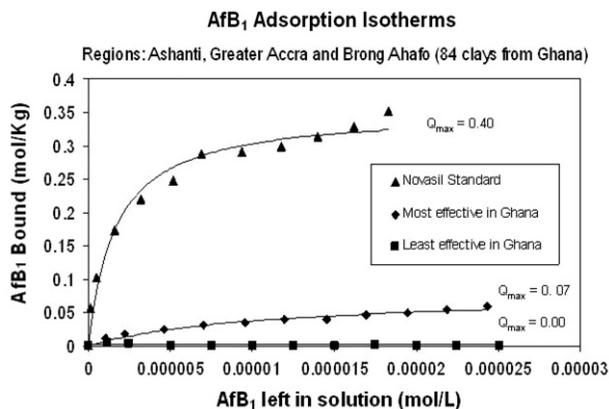


Figure 3. Representative isothermal plots of the most effective and least effective clay samples from Ghana for the sorption of AfB₁. These plots are compared with a standard isotherm for NovaSil. The most effective sample from Ghana was obtained from the Brong/Ahafo Region ($Q_{\max} = 0.07$), and the least effective sample was obtained from the Ashanti Region ($Q_{\max} = 0.00$). None was comparable with NovaSil ($Q_{\max} = 0.40$), and would not be expected to decrease the bioavailability of AfB₁.

locations in the country, we found that none of these sorbed aflatoxins with high affinity and capacity (Figure 3).

Phase IIa clinical intervention trial with NS in Ghana. Aflatoxin contamination in food products remains a serious burden in the developing world where a lack of untainted food supplies and poverty present a major and persistent challenge to many people in affected areas (McAlpin et al. 2002; Shephard 2003). Avoiding consumption of aflatoxin-contaminated foods is one of the most fundamental approaches for reducing risk of aflatoxicosis in humans. However, this is simply not feasible for many communities in developing countries and therefore emphasizes the need for viable intervention strategies to manage aflatoxin contaminated diets and treat aflatoxicosis. A recent study involved a 3-month double-blind and placebo controlled, phase IIa clinical trial conducted in the Ejura-Sekyedumase district, Ashanti Region, Ghana (Afriyie-Gyawu et al. 2007). The objective was to evaluate the safety, efficacy, and tolerance of dietary NS when administered to humans for the prevention of aflatoxin exposure and toxicity. The study protocol was approved by the Institutional Review Boards of Texas A&M University and its counterpart in Ghana for Ethical Clearance. Five hundred and seven volunteers were clinically screened to evaluate their general health, pregnancy status, and blood AfB₁-albumin adduct levels, and 177 of them were enrolled as study participants. Subjects were randomly assigned to three groups: high-dose (HD),



Figure 4. NS capsules for the phase IIa clinical intervention trial in Ghana. Treatment doses were 0, 1.5 or 3.0 g NS day⁻¹ before meals with water (microcrystalline cellulose was used as a placebo).

low-dose (LD) and placebo-control (PL) groups that received 3.0, 1.5 and 0 g NS day⁻¹, respectively, in capsules (Figure 4). To ensure compliance to treatment regimens and participant well-being, trained study monitors supervised administration of the encapsulated NS to participants and recorded side effects daily. On-site physicians performed physical examinations monthly. Blood and urine samples were collected for laboratory analysis. Over 90% of the participants completed the study, and compliance rate was more than 97%. Also, 99% of the time participants reported no side effects throughout the study. Mild to moderate adverse health events (approximately 0.5% of the time) were recorded in some participants but none of them appeared to be associated with NS treatment. No NS-related, significant differences were shown in haematology, liver and kidney functions, and electrolytes among the three groups. In the serum biochemical analysis, isolated statistical differences in a few parameters were detected but no trends of association or dose-dependency were observed, and were all within the normal physiological ranges (Afriyie-Gyawu et al. 2007).

This study represents the first phase IIa clinical intervention trial to evaluate the safety and efficacy of NS clay in human subjects. Results suggest that short-term inclusion of NS in the diet at a minimal effective dose (MED) of 0.25% (w/w) would not likely produce overt toxicity in humans. Moreover, the results of this study support the application of NS for the management of aflatoxicosis in humans who are acutely exposed to high levels of dietary aflatoxins. Additional results from this study indicate that ingestion of capsules containing an MED of NS, significantly reduce biomarkers of aflatoxin exposure

in the blood and urine from study participants (Wang et al. 2007). Further studies are planned to optimize the dosimetry and delivery methods for NS clay. Also, phase IIb, phase III intervention, and epidemiological studies are needed to confirm the safety and efficacy of NS for long-term therapy and the potential inclusion in foods for humans in areas at high risk for aflatoxicosis.

Summary

NS clay is commonly used as an anti-caking agent in animal feeds. Importantly, its inclusion in feed (at relatively low levels) may also serve to protect animals by tightly sorbing aflatoxins in the stomach and intestines resulting in decreased bioavailability. Based on numerous studies, it is anticipated that NS clay-based enterosorption of aflatoxins in animals will result in improved growth rates, feed conversions and general health along with diminished aflatoxin residues in foods of animal origin (such as milk).

Our recent findings from clinical intervention trials with NS are of particular relevance to populations in developing countries where the incidence of HCC and adverse health impacts from frequent exposures to aflatoxin are often elevated. Moreover, the use of NS for the protection of humans that are at high risk for HCC and aflatoxicosis appears to be culturally acceptable and sustainable. Eventually, the preferred delivery of NS may be through its inclusion in salt (like iodine), taking advantage of its anticaking properties, or as an additive in common groundnut and maize-based foods. Extensive research with NS clay and other sorbent materials suggests that potential mycotoxin enterosorbents (e.g. chemical and biological binders and/or sequestering agents) should be rigorously evaluated *in vitro* and *in vivo*. These should meet the following criteria:

- Favourable thermodynamic characteristics of sorption.
- Tolerable levels of priority metals, dioxins/furans and other hazardous substances.
- Safety and efficacy in multiple animal species.
- Safety and efficacy in long-term studies.
- Negligible interactions with vitamins, iron and zinc.

Acknowledgements

This work was supported by research grants from the US Agency for International Development (USAID LAG-G-00-96-90013-00) through Peanut CRSP of the University of Georgia and NIEHS P42-ES04917.

References

- Abdel-Wahhab M, Nada S, Farag I, Abbas N, Amra H. 1998. Potential protective effect of HSCAS and bentonite against dietary aflatoxicosis in rat: With special reference to chromosomal aberration. *Natural Toxins* 6:211.
- Afriyie-Gyawu E, Ankrah N-A, Huebner H, Ofosuhenne M, Kumi J, Johnson N, Tang L, Xu L, Jolly P, Ellis P, et al. 2007. NovaSil clay intervention in Ghanaians at high risk for aflatoxicosis, Part I: Study design and clinical outcomes. *Food Additives and Contaminants* (accepted).
- Afriyie-Gyawu E, Mackie J, Dash B, Wiles M, Taylor J, Huebner H, Tang L, Guan H, Wang JS, Phillips T. 2005. Chronic toxicological evaluation of dietary NovaSil clay in Sprague-Dawley rats. *Food Additives and Contaminants* 22:259.
- Aguilar F, Hussain S, Cerutti P. 1993. Aflatoxin B₁ induces the transversion of G → T in codon 249 of the p53 tumor suppressor gene in human hepatocytes. *Proceedings of the National Academy of Sciences, USA* 90: 8586.
- Asao T, Buechi G, Abdel-Kader MM, Chang SB, Wick EL, Wogan GN. 1963. Aflatoxins B and G. *Journal of the American Chemical Society* 85:1706–1707.
- Beaver R, Wilson D, James M, Haydon K. 1990. Distribution of aflatoxins in tissues of growing pigs fed an aflatoxin-contaminated diet amended with a high affinity aluminosilicate sorbent. *Veterinary and Human Toxicology* 32:16.
- Bonna R, Aulerich R, Bursian S, Poppenga R, Braselton W, Watson G. 1991. Efficacy of hydrated sodium calcium aluminosilicate and activated charcoal in reducing the toxicity of dietary aflatoxin to mink. *Archives of Environmental Contamination and Toxicology* 20:441.
- Breinholt V, Schimerlik M, Dashwood R, Bailey G. 1995. Mechanisms of chlorophyllin anticarcinogenesis against aflatoxin B₁: Complex formation with the carcinogen. *Chemical Research in Toxicology* 8:506.
- Bulatao-Jayme J, Almero E, Castro M, Jardeleza M, Salamat L. 1982. A case-control dietary study of primary liver cancer risk from aflatoxin exposure. *International Journal of Epidemiology* 11:112–119.
- Bursian S, Aulerich R, Cameron J, Ames N, Steficek B. 1992. Efficacy of hydrated sodium calcium aluminosilicate in reducing the toxicity of dietary zearalenone to mink. *Journal of Applied Toxicology* 12:85–90.
- Carretero MI. 2002. Clay minerals and their beneficial effects upon human health. *Applied Clay Science* 21:155–163.
- Carson M, Smith T. 1983. Role of bentonite in prevention of T-2 toxicosis in rats. *Journal of Animal Science* 57:1498.
- Centers for Disease Control and Prevention (CDC). 2004. Outbreak of aflatoxin poisoning — Eastern and Central Provinces, Kenya. *Morbidity and Mortality Weekly Report* 53: 790–793.
- Cerdchai R, Paisansarakit A, Khajarern J. 1990. Effect of hydrated sodium calcium aluminosilicate (NovaSil) on reducing aflatoxicosis in ducks. In *Proceedings of the 7th FAVA Congress, Pattaya*. p. 391.
- Chestnut A, Anderson P, Cochran M, Fribourg H, Gwinn K. 1992. Effects of hydrated sodium calcium aluminosilicate on fescue toxicosis and mineral absorption. *Journal of Animal Sciences* 70:2838.
- Chi WJ, Doong SL, Lin-Shiau SY, Boone CW, Kelloff GJ, Lin JK. 1998. Oltipraz, a novel inhibitor of hepatitis B virus transcription through elevation of p53 protein. *Carcinogenesis* 19:2133–2138.
- Chung T, Baker D. 1990. Phosphorus utilization in chicks fed hydrated sodium calcium aluminosilicate. *Journal of Animal Science* 68:1364.
- Chung T, Erdman Jr J, Baker D. 1990. Hydrated sodium calcium aluminosilicate: effects on zinc, manganese, vitamin A and riboflavin utilization. *Poultry Science* 69:1364.
- Colvin B, Sangster L, Hayden K, Bequer R, Wilson D. 1989. Effect of high affinity aluminosilicate sorbent on prevention of aflatoxicosis in growing pigs. *Veterinary and Human Toxicology* 31:46.
- Council for Agricultural Science and Technology (CAST). 1989. *Mycotoxins: economic and health risks*. Task Force Report No. 116. Ames, IA: CAST. p. 1–91.
- Dashwood R, Negishi T, Hayatsu H, Breinholt V, Hendricks J, Bailey G. 1998. Chemopreventive properties of chlorophylls towards aflatoxin B₁: A review of the antimutagenicity and anticarcinogenicity data in rainbow trout. *Mutation Research* 399:245.
- Davidson J, Babish J, Delaney K, Taylor D, Phillips TD. 1987. Hydrated sodium calcium aluminosilicate decreases the bioavailability of aflatoxin in the chicken. *Poultry Science* 66:89.
- Diamond J. 1999. *Evolutionary biology: Dirty eating for healthy living*. *Nature* 400:120.
- Dragsted LO, Strube M, Larsen JC. 1993. Cancer-protective factors in fruits and vegetables: Biochemical and biological background. *Pharmacology and Toxicology* 72(Suppl. 1): 116–135.
- Dwyer M, Kubena L, Harvey R, Mayura K, Sarr A, Buckley S, Bailey R, Phillips TD. 1997. Effects of inorganic adsorbents and cyclopiazonic acid in broiler chickens. *Poultry Science* 76:1141.
- Eaton D, Ramsdell H, Neal G. 1994. Biotransformation of aflatoxins. In: Eaton L, Groopman J, editors. *The toxicology of aflatoxins, human health, veterinary agricultural significance*. New York, NY: Academic Press.
- Egner P, Wang J, Zhu Y, Zhang B, Wu Y, Zhang Q, Qian G, Kuang S, Gange S, Jacobson L, et al. 2001. Chlorophyllin intervention reduces aflatoxin–DNA adducts in individuals at high risk for liver cancer. *Proceedings of the National Academy of Sciences, USA* 98: 14601.
- Ellis J, Harvey R, Kubena L, Bailey R, Clement B, Phillips TD. 1990. Reduction of aflatoxin M₁ residues in milk utilizing hydrated sodium calcium aluminosilicate. *Toxicologist* 10:163 (abst).
- Ellis R, Clements M, Tibbetts A, Winfree R. 2000. Reduction of the bioavailability of 20 µg/kg aflatoxin in trout feed containing clay. *Aquaculture* 183:179.
- Essigmann JM, Croy RG, Nadzan AM, Busby WF, Reinhold VN, Buchi GB, Wogan GN. 1977. Structural identification of the major DNA adduct formed by aflatoxin B₁ in vitro. *Proceedings of the National Academy of Sciences, USA* 74: 1870–1874.
- Fujiki H, Suganuma M, Imai K, Nakachi K. 2002. Green tea: Cancer preventive beverage and/or drug. *Cancer Letters* 188:9.
- Garland T, Reagor J. 2001. Chronic canine aflatoxicosis and management of an epidemic. In: deKoe W, Samson R, Van Egmond H, Gilbert J, Sabina M, editors. *Mycotoxins and phycotoxins in perspective at the turn of the millennium*. Wageningen: Ponsen & Looven. pp 231–236.
- Gong YY, Cardwell K, Hounsa A, Egal S, Turner PC, Hall AJ, Wild CP. 2002. Dietary aflatoxin exposure and impaired growth in young children from Benin and Togo: Cross sectional study. *British Medical Journal* 325: 20–21.
- Gong YY, Cardwell K, Hounsa A, Egal S, Turner PC, Sutcliffe AE, Hall AJ, Cardwell K, Wild CP. 2004. Postweaning exposure to aflatoxin results in impaired child growth: A longitudinal study in Benin, West Africa. *Environmental Health Perspectives* 112:1334–1338.

- Gopalakrishnan S, Harris TM, Stone MP. 1990. Intercalation of aflatoxin B₁ in two oligodeoxynucleotide adducts: Comparative ¹H NMR analysis of d(ATC^{AFB}GAT).d(ATCGAT) and d(AT^{AFB}GCAT)². *Biochemistry* 29:10438–10448.
- Grant P. 1998. Investigation of the mechanism of aflatoxin B₁ adsorption to clays and sorbents through the use of isothermal analysis. PhD dissertation, Texas A&M University, College Station, TX.
- Groopman J, Cain L, Kensler T. 1988. Aflatoxin exposure in human populations: Measurements and relationship to cancer. *CRC Critical Reviews in Toxicology* 19:113–145.
- Guengerich FP, Arneson KO, Williams KM, Deng Z, Harris TM. 2002a. Reaction of aflatoxin B₁ oxidation products with lysine. *Chemical Research in Toxicology* 15:780–793.
- Guengerich FP, Voehler M, Williams KM, Deng Z, Harris TM. 2002b. Structure of the aflatoxin B₁ dialdehyde adduct formed from reaction with methylamine. *Chemical Research in Toxicology* 15:793–798.
- Gupta R, Dubois R. 2001. Colorectal cancer prevention and treatment by inhibition of cyclooxygenase-2. *Nature Reviews Cancer* 1:11–21.
- Haderlein S, Weissmahr K, Schwarzenbach R. 1996. Specific adsorption of nitroaromatic explosives and pesticides to clay minerals. *Environmental Science and Technology* 30:612.
- Harvey R, Kubena L, Elissalde M, Corrier D, Phillips TD. 1994. Comparison of two hydrated sodium calcium aluminosilicate compounds to experimentally protect growing barrows from aflatoxicosis. *Journal of Veterinary Diagnostic Investigation* 6:88.
- Harvey R, Kubena L, Elissalde M, Phillips TD. 1993. Efficacy of zeolitic ore compounds on the toxicity of aflatoxin to growing broiler chickens. *Avian Diseases* 37:67.
- Harvey R, Kubena L, Phillips TD, Corrier D, Elissalde M, Huff W. 1991a. Diminution of aflatoxin toxicity to growing lambs by dietary supplementation with hydrated sodium calcium aluminosilicate. *American Journal of Veterinary Research* 52:152.
- Harvey R, Phillips TD, Ellis J, Kubena L, Huff W, Petersen D. 1991b. Effects of aflatoxin M₁ residues in milk by addition of hydrated sodium calcium aluminosilicate to aflatoxin-contaminated diets of dairy cows. *American Journal of Veterinary Research* 52:1556.
- Hayatsu H, Arimoto S, Negishi T. 1988. Dietary inhibitors of mutagenesis and carcinogenesis. *Mutation Research* 202:429–446.
- Hesseltine CW. 1979. Some important fermented foods of Mid-Asia, the Middle East, and Africa. *Journal of the American Oil Chemists' Society* 56:367–374.
- Hinton DM, Myers MJ, Raybourne RA, Francke-Carroll S, Sotomayor RE, Shaddock J, Warbritton A, Chou MW. 2003. Immunotoxicity of aflatoxins in rats: Effects on lymphocytes and the inflammatory response in a chronic intermittent dosing study. *Toxicological Sciences* 73:362–377.
- Huebner H, Herrera P, Phillips TD. 2004. Clay-based interventions for the control of chemical and microbial hazards in food and water. In: Beier R, Pallai S, Phillips TD, editors. *Preharvest and postharvest food safety: Contemporary issues and future directions*. Ames, IA: IFT Press. pp 389–402.
- Huebner H, Lemke S, Ottinger S, Mayura K, Phillips TD. 1999. Molecular characterization of high affinity, high capacity clays for the equilibrium sorption of ergotamine. *Food Additives and Contaminants* 16:159.
- Huebner H, Phillips TD. 2003. Clay-based affinity probes for elective cleanup and determination of Aflatoxin B₁ Using Nanostructured Montmorillonite on Quartz. *Journal of AOAC International* 86:534.
- Huff W, Kubena L, Harvey R, Phillips TD. 1992. Efficacy of a hydrated sodium calcium aluminosilicate to reduce the individual and combined toxicity of aflatoxin and ochratoxin A. *Poultry Science* 71:64.
- International Agency for Research on Cancer (IARC). 1976. Some naturally occurring substances. Monographs on the evaluation of the carcinogenic risk of chemicals to man, Vol. 10. Lyon: IARC.
- International Agency for Research on Cancer (IARC). 1987. Aflatoxins. Monograph on the evaluation of carcinogenic risk to humans, Suppl. 7. Lyon: IARC.
- International Agency for Research on Cancer (IARC). 1993. Aflatoxins. Monograph on the evaluation of carcinogenic risk to humans, Suppl. 7. Lyon: IARC.
- Jayaprakash M, Gowda R, Vijayasarithi S, Seshadri S. 1992. Adsorbent efficacy of hydrated sodium calcium aluminosilicate in induced aflatoxicosis in broilers. *Indian Journal of Veterinary Pathology* 16:102.
- Jiang Y, Jolly P, Ellis W, Wang J, Phillips T, Williams J. 2005. Aflatoxin B₁ albumin adduct levels and cellular immune status in Ghanaians. *International Immunology* 17:807–814.
- Johns T, Duquette M. 1991. Detoxification and mineral supplementation as functions of geophagy. *American Journal of Clinical Nutrition* 53:448.
- Kensler TW, Groopman JD, Sutter TR, Curphey TJ, Roebuck BD. 1999. Development of cancer chemopreventive agents: oltipraz as a paradigm. *Chemical Research in Toxicology* 12:113–126.
- Klaunig J, Kamendulis L. 1999. Mechanisms of cancer chemoprevention in hepatic carcinogenesis: Modulation of focal lesion growth in mice. *Toxicological Sciences* 52:101.
- Kubena L, Harvey R, Huff W, Corrier D, Phillips TD. 1990b. Ameliorating properties of a hydrated sodium calcium aluminosilicate on the toxicity of aflatoxin and T-2 toxin. *Poultry Science* 69:1078.
- Kubena L, Harvey R, Huff W, Yersin A, Elissalde M, Witzel D. 1993. Efficacy of a hydrated sodium calcium aluminosilicate to reduce the toxicity of aflatoxin and diacetoxyscirpenol. *Poultry Science* 72:51.
- Kubena L, Harvey R, Phillips TD, Corrier D, Huff W. 1990a. Diminution of aflatoxicosis in growing chickens by dietary addition of a hydrated sodium calcium aluminosilicate. *Poultry Science* 69:727.
- Kubena L, Huff W, Harvey R, Yersin A, Elissalde M, Witzel D, Giroir L, Phillips TD. 1991. Effects of hydrated sodium calcium aluminosilicate on growing turkey poults during aflatoxicosis. *Poultry Science* 70:1823.
- Lancaster M, Jenkins F, Philp J. 1961. Toxicity associated with certain samples of groundnuts. *Nature* 192:1095.
- Lang SS. 2006. Dogs keep dying: Too many owners unaware of toxic dog food. *Cornell University ChronicleOnline*, 6 January (available at: <http://www.news.cornell.edu/stories/Jan06/dogs.dying.ssl.html>).
- Ledoux D, Rottinhaus G, Bermudez A, Alonso-Debolt M. 1999. Efficacy of a hydrated sodium calcium aluminosilicate to ameliorate the toxic effects of aflatoxin in broiler chicks. *Poultry Science* 78:204.
- Leeson S, Diaz G, Summers J. 1995. In: *Poultry metabolic disorders and mycotoxins*. Guelph: University Books.
- Lemke S. 2000. Investigation of clay-based strategies for the protection of animals from the toxic effects of selected mycotoxins. PhD dissertation, Texas A&M University, College Station, TX.
- Lewis L, Onsongo M, Njapau H, Schurz-Rogers H, Lubber G, Kieszak S, Nyamongo J, Backer L, Dahiye AM, Misore A, et al. 2005. Aflatoxin contamination of commercial maize products during an outbreak of acute aflatoxicosis in eastern and central Kenya. *Environmental Health Perspectives* 113:1763–1767.

- Lindemann M, Blodgett D, Kornegay E, Schurig G. 1993. Potential ameliorators of aflatoxicosis in weaning/growing swine. *Journal of Animal Science* 71:171.
- Marquez R, Hernandez I. 1995. Aflatoxin adsorbent capacity of two Mexican aluminosilicates in experimentally contaminated chick diets. *Food Additives and Contaminants* 12:431.
- Masimango N, Remacle J, Ramaut J. 1979. Elimination, par des argiles gonflantes, de L'aflatoxine B₁ des milieux contaminés. *Annales de la Nutrition et de l'Alimentation* 33:137.
- Mayura K, Abdel-Wahhab M, McKenzie K, Sarr A, Edwards J, Naguib K, Phillips TD. 1998. Prevention of maternal and developmental toxicity in rats via dietary inclusion of common aflatoxin sorbents: Potential for hidden risks. *Toxicological Sciences* 41:175.
- McAlpin CE, Wicklow DT, Horn BW. 2002. DNA fingerprinting analysis of vegetative compatibility groups in *Aspergillus flavus* from a peanut field in Georgia. *Plant Diseases* 86:254.
- McMahon G, Davis E, Wogan GN. 1987. Characterization of c-Ki-ras oncogene alleles by direct sequencing of enzymically amplified DNA from carcinogen-induced tumors. *Proceedings of the National Academy of Sciences, USA* 84: 4974-4978.
- McMahon G, Hanson L, Lee JJ, Wogan GN. 1986. Identification of an activated c-Ki-ras oncogene in rat liver tumors induced by aflatoxin B₁. *Proceedings of the National Academy of Sciences, USA* 83: 9418-9422.
- Miller DM, Stuart SP, Crowell WA, Cole RJ, Goven AJ, Brown J. 1978. Aflatoxicosis in swine: Its effect on immunity and relationship to salmonellosis. *Proceedings of the Annual Meeting of the American Association of Veterinary Lab Diagnostics* 21:135-146.
- Muller R, Carlson C, Semeniuk G, Harshfield G. 1970. Response of chicks, ducklings, goslings, pheasants, and poult to graded levels of aflatoxins. *Poultry Science* 49:1346.
- Nahm K. 1995. Prevention of aflatoxicosis by addition of antioxidants and hydrated sodium calcium aluminosilicate to the diet of young chicks. *Japanese Journal of Poultry Science* 32:117.
- Nakachi K, Matsuyama S, Miyake S, Sugauma M, Imai K. 2000. Preventive effects of drinking green tea on cancer and cardiovascular disease: Epidemiological evidence for multiple targeting prevention. *BioFactors* 13:49.
- Patterson R, Young L. 1993. Efficacy of hydrated sodium calcium aluminosilicate, screening and dilution in reducing the effects of mold contaminated corn in pigs. *Canadian Journal of Animal Science* 73:616.
- Peraica M, Radic B, Lucic A, Pavlovic M. 1999. Toxic effects of mycotoxins in humans. *Bulletin of the World Health Organization* 77:754-66.
- Peska J, Bondy G. 1994. Immunotoxic effects of mycotoxins. In: Miller J, Trenholm H, editors. *Mycotoxins in grain: Compounds other than aflatoxin*. St Paul, MN: Eagan.
- Phillips TD. 1999. Dietary clay in the chemoprevention of aflatoxin-induced disease. *Toxicology Sciences* 52:118.
- Phillips TD, Afriyie-Gyawu E, Wang JS, Williams J, Huebner H. 2006. The potential of aflatoxin sequestering clay. In: Barug D, Bhatnagar D, Van Egmond H, Van der Kamp J, Van Osenbruggen W, Visconti A, editors. *The mycotoxin fact book*. Wageningen: Wageningen Academic Publ. pp 329-346.
- Phillips TD, Clement B, Kubena L, Harvey R. 1990. Detection and detoxification of aflatoxins: Prevention of aflatoxicosis and aflatoxin residues with hydrated sodium calcium aluminosilicates. *Veterinary and Human Toxicology* 32:15.
- Phillips TD, Clement B, Park D. 1994. Approaches to reduction of aflatoxin. In: Eaton L, Groopman J, editors. *The toxicology of aflatoxins, human health, veterinary agricultural significance*. New York, NY: Academic Press.
- Phillips TD, Kubena L, Harvey R, Taylor D, Heidelbaugh N. 1987. Mycotoxin hazards in agriculture: new approach to control. *Journal of American Veterinary Medical Association* 190:1617 (abst).
- Phillips TD, Kubena L, Harvey R, Taylor D, Heidelbaugh N. 1988. Hydrated sodium calcium aluminosilicate: A high affinity sorbent for aflatoxin. *Poultry Science* 67:243.
- Phillips TD, Lemke SL, Grant PG. 2002. Characterization of clay-based enterosorbents for the prevention of aflatoxicosis. *Advances in Experimental Medicine and Biology* 504:157-171.
- Phillips TD, Sarr A, Grant P. 1995. Selective chemisorption and detoxification of aflatoxins by phyllosilicate clay. *Natural Toxins* 3:204.
- Phillips TD, Sarr AB, Clement B, Kubena L, Harvey R. 1991. Prevention of aflatoxicosis in farm animals via selective chemisorption of aflatoxin. In: Bray G, Ryan D, editors. *Mycotoxins, cancer and health*. Vol. 1. Baton Rouge, LA: Louisiana State University Press.
- Pimpukdee K, Kubena LF, Bailey CA, Huebner HJ, Afriyie-Gyawu E, Phillips TD. 2004. Aflatoxin-induced toxicity and depletion of hepatic vitamin A in young broiler chicks: Protection of chicks in the presence of low levels of NovaSil PLUS in the diet. *Poultry Science* 83:737.
- Qin G, Gopalan-Kriczky P, Su J, Ning Y, Lotlikar P. 1997. Inhibition of aflatoxin B₁-induced initiation of hepatocarcinogenesis in the rat by green tea. *Cancer Letters* 112:149.
- Ramsbottom J. 1953. *Mushrooms and toadstools*. London: Collins.
- Richard JL, Thurston JR, Pier AC. 1978. Effects of mycotoxins on immunity. *Toxin*. In: Rosenberg P, editor. *Animal, plant and microbial*. New York, NY: Pergamon. pp 801-817.
- Rodricks JV, Stoloff L. 1977. Aflatoxin residues from contaminated feed in edible tissues of food-producing animals. In: *Mycotoxins in Human and Animal Health, Proceedings of a Conference*. pp 67-79.
- Sabbioni G, Skipper PL, Büchi G, Tannenbaum SR. 1987. Isolation and characterization of the major serum albumin adduct formed by aflatoxin B₁ in vivo in rats. *Carcinogenesis* 8:819-824.
- Sargeant K, O'Kelly J, Carnaghan R, Allcroft R. 1961. The assay of a toxic principle in certain groundnut meals. *Veterinary Record* 73:1219.
- Sarr A, Mayura K, Kubena L, Harvey R, Phillips TD. 1995. Effects of phyllosilicate clay on the metabolic profile of aflatoxin B₁ in Fischer-344 rats. *Toxicology Letters* 75:145.
- Shepard G. 2003. Aflatoxin and food safety: recent African perspectives. *Journal of Toxicology, Toxin Reviews* 22: 267-286.
- Siller W, Ostler D. 1961. The histopathology of an enterohepatic syndrome of turkey poults. *Veterinary Record* 73:134.
- Smith E, Phillips TD, Ellis J, Harvey R, Kubena L, Thompson J, Newton G. 1994. Dietary hydrated sodium calcium aluminosilicate reduction of aflatoxin M₁ residue in dairy goat milk and effects on milk production and components. *Journal of Animal Science*, 72:677.
- Smith T. 1984. Spent canola oil bleaching clays: Potential for treatment of T-2 toxicosis in rats and short-term inclusion in diets of immature swine. *Canadian Journal of Animal Science* 64:725.
- Sporn M, Suh N. 2002. Chemoprevention: An essential approach to controlling cancer. *Nature Reviews Cancer* 2:537-543.

- Turner PC, Collinson AC, Cheung YB, Gong YY, Hall AJ, Prentice AM, Wild CP. 2007. Aflatoxin exposure in utero causes growth faltering in Gambian infants. *International Journal of Epidemiology* Advance Access (18 June).
- Turner PC, Moore SE, Hall AJ, Prentice AM, Wild CP. 2003. Modification of immune function through exposure to dietary aflatoxin in Gambian children. *Environmental Health Perspectives* 111:217–220.
- Turner PC, Sylla A, Gong YY, Diallo MS, Sutcliffe AE, Hall AJ, Wild CP. 2005. Reduction in exposure to carcinogenic aflatoxins by postharvest intervention measures in West Africa: A community-based intervention study. *Lancet* 365:1950–1956.
- Van Rensburg SJ, Altenkirk B. 1974. *Claviceps purpurea* — ergotism. In: IFH, editor. *Mycotoxins purchase*. Elsevier. pp 69–96.
- Voss K, Dorner J, Cole R. 1993. Amelioration of aflatoxicosis in rats by volclay NF-BC, microfine bentonite. *Journal of Food Protection* 56:595.
- Wang JS, Luo H, Billam M, Wang Z, Guan H, Tang L, Goldston T, Afriyie-Gyawu E, Lovett C, Griswold J, et al. 2005. Short-term safety evaluation of processed calcium montmorillonite clay (NovaSil) in humans. *Food Additives and Contaminants* 22:270–279.
- Wang JS, Qian GS, Zarba A, He X, Zhu YR, Zhang BC, Jacobson L, Gange SJ, Munoz A. 1996. Temporal patterns of aflatoxin–albumin adducts in hepatitis B surface antigen-positive and antigen-negative residents of Daxin, Qidong County, People’s Republic of China. *Cancer Epidemiology, Biomarkers and Prevention* 5:253–261.
- Wang P, Afriyie-Gyawu E, Tang Y, Johnson NM, Xu L, Tang L, Huebner HJ, Ankrah NA, Ofori-Adjei D, Ellis W, et al. 2007. NovaSil clay intervention in Ghanaians at high risk for aflatoxicosis: II. Reduction in biomarkers of aflatoxin in blood and urine. *Food Additives and Contaminants* (in press).
- Wang JS, Shen X, He X, Zhu YR, Zhang BC, Wang JB, Qian GS, Kuang SY, Zarba A, Egner PA, et al. 1999. Protective alterations in phase 1 and 2 metabolism of aflatoxin B1 by oltipraz in residents of Qidong, People’s Republic of China. *Journal of the National Cancer Institute* 91:347–354.
- Wild CP, Hall AJ. 2000. Primary prevention of hepatocellular carcinoma in developing countries. *Mutation Research* 462:381–393.
- Wild CP, Turner PC. 2002. The toxicology of aflatoxins as a basis for public health decisions. *Mutagenesis* 17:471–481.
- Wiles M, Huebner H, Afriyie-Gyawu E, Taylor R, Bratton G, Phillips TD. 2004. Toxicological evaluation and metal bio-availability in pregnant rats following exposure to clay minerals in the diet. *Journal of Toxicology and Environmental Health: Part A* 67:863–874.
- Williams JH, Phillips TD, Jolly PE, Stiles JK, Jolly CM, Aggarwal D. 2004. Human aflatoxicosis in developing countries: A reviews of toxicology, exposure, potential health consequences, and interventions. *American Journal of Clinical Nutrition* 80:1106–1122.
- Wogan GN. 1992. Molecular epidemiology in cancer risk assessment and prevention: Recent progress and avenues for future research. *Environmental Health Perspectives* 11:47–54.
- World Health Organization (WHO). 1998. Safety evaluation of certain food activities and contaminants. WHO Food Additives Series No. 40 Prepared by the Forty-ninth Meeting of the Joint FAO/WHO Expert Committee on Food Additives. Geneva: WHO, International Programme on Chemical Safety.